

Penicillin Biosynthesis: Active Substrates derived by Methoxy Substitution in the Valinyl Residue of the Natural Substrate

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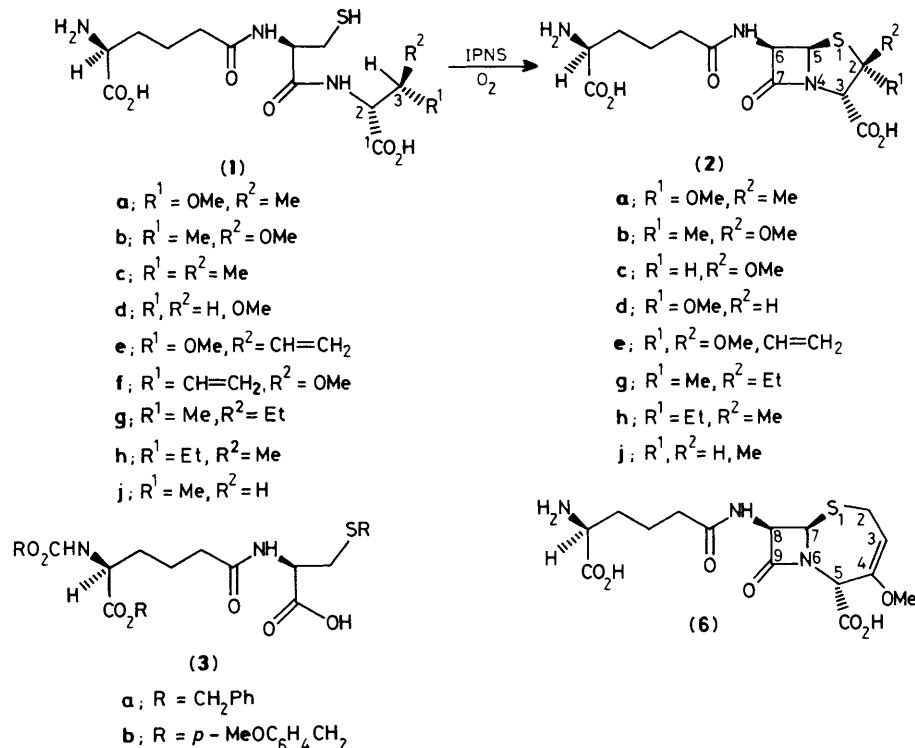
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The structure–reactivity profile of tripeptides modified by methoxy substitution in the valinyl moiety of L-(α -aminoadipoyl)-L-cysteinyl-D-valine with the enzyme isopenicillin N synthase has been examined; substrate bulk and absolute configuration at the oxygen-substituted carbon were found to play crucial roles in determining substrate reactivity.

Recently we described that a new antibacterial penicillin containing a 2 α -methoxy group (**2a**) was obtained by enzymatic synthesis from the tripeptide δ -(L- α -aminoadipoyl)-L-cysteinyl-D-(*O*-methyl-allothreonine) (**1a**) with isopenicillin N synthase (IPNS) from *Cephalosporium acremonium* CO 728.¹ The absolute configuration at C-3 of so-modified peptides was found to be an important factor since the *O*-methyl threonyl tripeptide (**1b**) was not a β -lactam producing substrate with IPNS, *i.e.*, (**1b**) \nrightarrow (**2b**). In order to evaluate the generality of methoxy substitution for methyl in the valine moiety of the natural substrate (**1c**), we have synthesised and tested a series of methoxy-modified tripeptides.

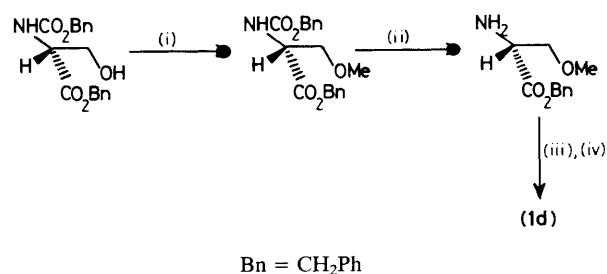
Initially the *O*-methylserinyl tripeptide (**1d**) was synthesised following standard peptide coupling and deprotection procedures² (Scheme 1). Incubation of (**1d**) with purified IPNS under the usual conditions³ gave after protein precipitation two new β -lactam products. Purification by h.p.l.c. [reverse phase octadecylsilane column, i, 25 mM NH₄HCO₃ as eluant; ii, 0.05% HCO₂H in H₂O as eluant] gave firstly the 2 β -methoxypenam (**2c**), δ_{H} (500 MHz, D₂O)[†] 1.65–1.95 (4H, 2 \times m, CH₂CH₂CH₂CO), 2.38–2.45 (2H, m, CH₂CO),

[†] ¹H N.m.r. spectra were referenced to sodium 3-trimethylsilyl-tetradecuteriopropionate = 0.00 p.p.m.



3.32 (3H, s, OMe), 3.70–3.75 [1H, m, $\text{CH}(\text{CH}_2)_3$], 4.98 (1H, s, 3-H), 5.38, 5.48 (2H, ABq, J 4 Hz, 5,6-H), 5.63 (1H, s, 2-H); m/z (positive argon fast atom bombardment) 362 (MH^+), which showed no detectable antibacterial activity towards *Staphylococcus aureus* N.C.T.C. 6571 or *Micrococcus luteus* DS 292 at a concentration of $50 \mu\text{g ml}^{-1}$,[‡] and secondly [h.p.l.c. system i, only] the 2 α -methoxypenam (2d) δ_{H} (500 MHz, D_2O)[†] 1.63–1.93 (4H, $2 \times$ m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{CO}$), 2.35–2.40 (2H, m, CH_2CO), 3.36 (3H, s, OMe), 3.68–3.73 [1H, m, $\text{CH}(\text{CH}_2)_3$], 5.47, 5.59 (2H, ABq, J 4 Hz, 5,6-H), 5.84 (1H, d, J 6 Hz, 2-H), (3-H obscured), m/z (positive argon fast atom bombardment) 362 (MH^+), which showed antibacterial activity towards *S. aureus* N.C.T.C. 6571 and *M. luteus* DS 292 at a concentration of $50 \mu\text{g ml}^{-1}$.[‡] The stereochemistries of (2c) and (2d) as the 2 β - and 2 α -methoxy penams respectively follow from the observed couplings $J(\text{H}-2, \text{H}-3)$ of ca. 0 and 6 Hz respectively, and by comparison with literature coupling constant values.⁴

Secondly we studied the effect of methoxy substitution at C-3 of unsaturated tripeptides, via synthesis of the tripeptides (1e) and (1f) (Scheme 2). Incubation of (1e) with purified IPNS³ gave, after protein precipitation, two new β -lactam products. Purification by h.p.l.c. (reverse phase octadecylsilane column, 10 mM NH_4HCO_3 as eluant) gave firstly the 2-methoxy-2-vinyl penam (2e), δ_{H} (500 MHz, D_2O)[†] 1.63–1.72, 1.82–1.92 (4H, $2 \times$ m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{CO}$), 2.37–2.43 (2H, m, CH_2CO), 5.24, 5.67 (2H, ABq, J 4 Hz, 5,6-H), 5.42 (1H, d, J 11 Hz, $\text{CH}=\text{CH}_2$), 5.52 (1H, d, J 17 Hz, $\text{CH}=\text{CH}_2$), 5.88 (1H, dd, J 17, 11 Hz, $\text{CH}=\text{CH}_2$), [3-H, $\text{MeOCH}(\text{CH}_2)_3$ obscured], as a very minor product which gave antibacterial activity towards *Staphylococcus aureus* N.C.T.C. 6571 at a concentration of $25 \mu\text{g ml}^{-1}$,[‡] and secondly the 4-methoxy-homoceph-3-em (6),[§] ν_{max} (CaF_2 cells, D_2O) 1740s ($\text{C}=\text{O}$); δ_{H}



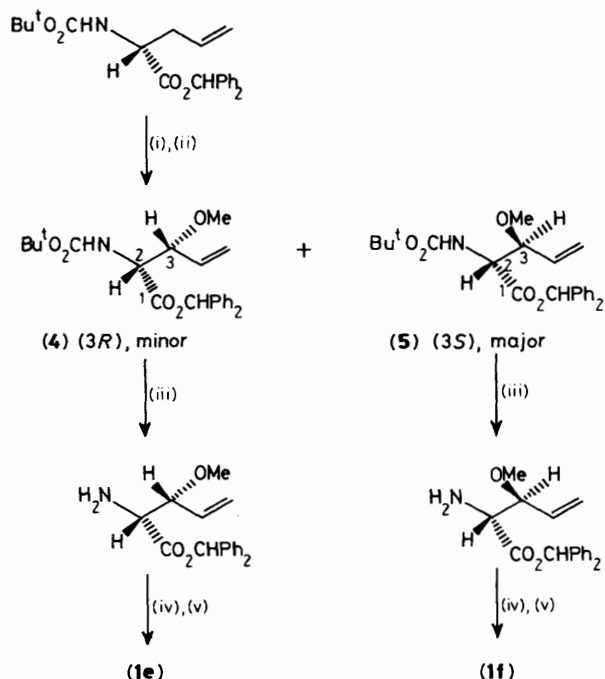
Scheme 1. Reagents: i, $\text{BF}_3 \cdot \text{OEt}_2$, CH_2N_2 ; ii, HBr/HOAc , then NEt_3 ; iii, coupling to (3a), ref. 2; iv, Na/NH_3 , ref. 2.

(500 MHz, D_2O)[†] 1.62–1.92 (4H, $2 \times$ m, $\text{CH}_2\text{CH}_2\text{CH}_2\text{CO}$), 2.34–2.42 (2H, m, CH_2CO), 3.31 (2H, ca. d, J 7.5 Hz, 2-H), 3.52 (3H, s, OMe), 3.65–3.72 [1H, m, $\text{CH}(\text{CH}_2)_3$], 4.93 (1H, s, 5-H), 5.13 (1H, ca. t, J 7.5 Hz, 3-H), 5.35, 5.58 (2H, ABq, J 4 Hz, 7,8-H), as a major product which showed no detectable antibacterial activity towards *S. aureus* N.C.T.C. 6571 at a concentration of $100 \mu\text{g ml}^{-1}$,[‡] m/z (positive argon fast atom bombardment) 388 (MH^+). The structure as (6) was consistent with selective ^1H n.m.r. spectroscopy decoupling experiments since irradiation of 2-H (δ_{H} 3.31) caused collapse of 3-H (δ_{H} 5.13) from a triplet to a singlet, and irradiation of 3-H caused collapse of 2-H from a doublet to a singlet. Incubation of the diastereoisomeric tripeptide (1f) with IPNS gave no detectable conversion to β -lactam products.

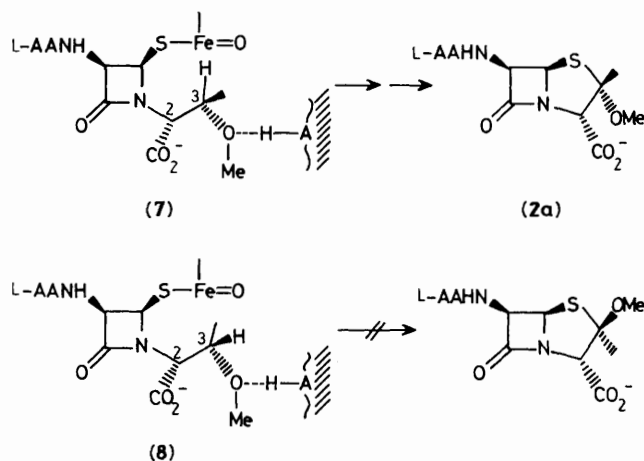
In summary these results and others^{1,7} demonstrate that if methoxy substitution at C-3 within the valinyl equivalent of the tripeptide provides the *allo* configuration, then such tripeptides can act as substrates for IPNS, whereas the corresponding diastereoisomeric *threo* configured tripeptides do not. In sharp contrast the tripeptides containing isoleucine, (1g) and *allo*-isoleucine, (1h), both cyclise to penicillins, (2g) and (2h) respectively, with retention of configuration in the

[‡] 100 μl of this solution was used for bioassay analysis by the 'holed plate' assay method.

[§] For (6) an α -configuration of the carboxy group is assumed, not proven.



Scheme 2. Reagents: i, SeO_2 , $\text{Bu}^t\text{O}_2\text{H}$, $\text{C}_2\text{H}_4\text{Cl}_2$, ref. 5; ii, TlOEt , MeI , dimethylformamide; iii, toluene-4-sulphonic acid, then NaHCO_3 ; iv, coupling to (3b), ref. 6; v, $\text{CF}_3\text{CO}_2\text{H}$, anisole, ref. 6.



Scheme 3. L-AA = L-α-aminoadipoyl.

carbon-sulphur bond-forming step.⁸ Since an ethyl group must be approximately isosteric with a methoxy group the difference between the two series must derive from some property of the methoxy group. One possibility is that a hydrogen bond between the methoxy group and an active site group restricts rotation around C(2)–C(3) in the intermediate,^{7,9} as in Scheme 3. Thus, the species (7) derived from the *allo*-threonine peptide can be cyclised, but that derived from the threonine, (8), is restrained in a conformation in which the β-hydrogen atom cannot be attacked by the active species. The formation of two epimeric methoxy penams, (2c) and (2d), from the *O*-methyl serine peptide (1d) may be compared with the α-aminobutyrate peptide (1j) which gave both epimeric monomethylpenams (2j), presumably through a rapidly rotating free radical,^{4,10} a process which is hindered by the presence of two substituents at the radical centre.

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